Isolation and Characterization of Atropisomers of Seven-Membered-Ring Benzolactams

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Supporting Information

ABSTRACT: The atropisomeric properties of seven-membered-ring benzolactams (7a-c and 8a) [1,5-benzodiazepin-2-one (a), 1,5-benzothiazepin-4-one (b), and 1-benzazepin-2one (c)] were examined. The atropisomers were isolated as the diastereomers with an (S)-phenethylamide moiety, which were characterized by X-ray crystallography, and the barriers to their interconversion were clarified.



INTRODUCTION

Seven-membered-ring benzolactams (I) (Figure 1) have been used as the core structures of various biologically active molecules. Lofendazam (I: X = NH),¹ diltiazem (I: X = S)² and benazepril (I: $X = CH_2$)³ are the typical therapeutic agents developed using these structures. Thus far, the conformation of these heterocycles has been extensively investigated to gain insight into the structures of these relatively flexible heterocycles.⁴ In essence, the important point is that compound I adopts chiral conformations because of an sp^2-sp^2 axis at Ar-N(C=O) to form the atropisomeric structures A and B (Figure 1)⁵ and the target molecules (receptors or enzymes) may recognize A and B for biological activity. However, there has been little discussion about the conformation from the viewpoint of the atropisomerism.⁶

In our previous papers,⁷ we described the atropisomeric properties of dibenzo [b,d] azepin-6-one (II: $\mathbf{a}-\mathbf{c}$), which is the core structure of a γ -secretase inhibitor, LY-411575⁸ (Figure 2). Compound II (a-c) was shown to exist as a racemate of the atropisomers II-A and II-B with an activation free-energy barrier to rotation (ΔG^{\dagger}) of 98–122 kJ/mol. X-ray analysis of the enantiomers of IIc revealed the presence of two $sp^2 - sp^2$ axes of Ar-Ar and Ar-N(C=O), although they exist only as a pair of enantiomers $[\mathbf{A}(a^1R,a^2S) \text{ and } \mathbf{B}(a^1S,a^2R)]$ without the presence of diastereomers $[(a^1R,a^2R) \text{ and } (a^1S,a^2S)]$. We assumed that this phenomenon occurred because the two axes move together like a gear. However, since the axial chirality at the aryl-amide bond has not been studied thoroughly⁹ compared with the biphenyl chirality, this is often regarded merely as a consequence of the biphenyl chirality and may not be fully confirmed. Herein we report the conformation of the seven-membered-ring benzolactams (I) to reveal and confirm the atropisomerism occurring at the Ar-N(C=O) axis.



Figure 1. Seven-membered-ring benzolactams (I) and atropisomeric structures A and B.



Figure 2. Conformation and thermodynamic stability of the atropisomers of dibenzo[*b*,*d*]azepin-6-one (IIa-c: A, B).

RESULTS AND DISCUSSION

Synthesis of Seven-Membered-Ring Benzolactams. The seven-membered-ring benzolactams (I) prepared are the 1,5benzodiazepin-2-ones (a series: $X = NCH_3$), 1,5-benzothiazepin-4-ones (**b** series: X = S), and 1-benzazepin-2-ones (**c** series: X =CH₂). The NH-lactams [1a-c (Y = H), 2a-c (Y = Cl)] were prepared from the corresponding benzene precursors with

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Figure 3. 1 H NMR (400 MHz, CDCl₃) spectra of 3a (a), 4a-COOH (b), 5a-COOH (c), and 6a-COOH (d).

NHCH₃, SH, and CH₂ moieties, respectively, as shown in Scheme 1. The compounds without a substituent at the ortho position on the benzene ring (Y = H) (1a-c) are the known compounds, 10-12 and compounds 2a-c, which have a chloro substituent at the ortho position (Y = Cl), were prepared according to the modified methods reported for the synthesis of 1a-c. It should be noted that the Beckmann rearrangement of 8-chloro-3,4-dihydronaphthalen-1-(2H)-one oxime with polyphosphoric acid (PPA) gave 9-chloro-1,3,4,5-tetrahydro-2H-1benzazepin-2-one (2c) and its regioisomer (1-one isomer) in a ratio of 53:47, whose stereochemistry was determined by the ¹H NMR (chemical shift of the C3-methylene protons: δ 2.38 (t, *J* = 7.2 Hz, 2H) for 2c and 3.11 (br, 2H) for the isomer). Compounds 3a-c with a phenyl group on the benzene ring were prepared in good yields in the Suzuki–Miyaura reaction of 2a-c. The benzolactams with N-substituents $[R = CH_2CO_2CH_3]$ (4-6) and $CH_2CONH(S)CH(CH_3)Ph(7, 8)$ were prepared from the corresponding NH-lactams (1-3) using conventional

methods, i.e., *N*-alkylation with methyl bromoacetate followed by hydrolysis and amidation with (*S*)-phenethylamine (Scheme 1). The intermediary compounds 4-6 are useful for preparation of the biologically active molecules. The orthosubstituent (Y = Cl, Ph) is introduced in the nuclei to provide a higher rotation barrier for the axis, which may result in freezing the conformation of the molecules.

Conformation of Seven-Membered-Ring Benzolactams (1-6). First, the conformations of compounds 1-6 were examined. The methylene protons of all the NH-lactams 1-3(a-c) showed sharp signals with AA'XX'-type splitting patterns in the ¹H NMR spectra, reflecting the rapid ring-flipping including the axis at aryl-NH(CO) (Figure 3a for 3a). On the other hand, compounds 4-6 which bear the N-methoxycarbonylmethyl moiety showed a different pattern in the ¹H NMR spectra. In compounds 4a-c, which are unsubstituted on the benzene ring (Y = H), the methylene protons of the lactam ring were observed as broad peaks without separation of the peaks of the geminal protons, and the methylene protons of the Nsubstituent also appeared as a broad signal (Figure 3b for 4a-COOH). Attempted separation of the atropisomers of 4a-c at room temperature using chiral HPLC failed. These results suggest that the axial chirality caused by the Ar-N(C=O) (sp^2-sp^2) axis is present, though the ring flipping occurs rather fast at room temperature.¹³ In contrast, in the ¹H NMR spectra compounds of 5a-c (Y = Cl) and 6a-c (Y = Ph), all the methylene protons of the lactam ring were observed as separated sharp peaks (ABXY-type splitting) and the methylene protons of the N-substituent as AB-quartet peaks (Figure 3c for 5a-COOH and Figure 3d for 6a-COOH), which indicate that the rotation around the Ar-N(C=O) axis is restricted by the chloro and phenyl groups to separate the protons as diastereotopic ones. The upper field shift of the AB-quartet peaks in 6a-COOH compared to those of 5a-COOH is obviously ascribed to the effect of the C9phenyl ring in 6a-COOH. As was expected from the ¹H NMR spectra, the atropisomers of 5a-c and 6a-c could be separated at room temperature using chiral HPLC. Compound **5b** (X = S, Y = Cl) was subjected to X-ray structure analysis, which revealed that the lactam ring takes a boatlike conformation and 5b exists as a racemate in the crystals; the aR and aS^{14} atropisomers are present in a unit cell (space group $P2_1/n$) (Figure 4).



Figure 4. X-ray crystal structure of **5b** (racemate): (a*R*)-form (left) and (a*S*)-form (right) in a unit cell.



Figure 5. X-ray crystal structures of 7a-A (aR, S) (left) and 7a-B (aS, S) (right).

Isolation and Structure Analysis of Atropisomers of Seven-Membered-Ring Benzolactams (7a–c and 8a). For isolation and characterization of the atropisomers, (S)-phenethylamide derivatives of 5a-c and 6a (7a–c and 8a) (Y = Cl and Ph) were prepared to separate the atropisomers as diastereomers (Scheme 1).

Compound 7a (X = NCH₃, Y = Cl) was obtained from 5a via the carboxylic acid as a mixture of the diastereomers (A and B) (80% yield) in a ratio of 1:0.7 (by ¹H NMR), which were separated on column chromatography using SiO₂ to afford 7a-A and 7a-B as crystals. Investigation of the reaction (amidation) using HPLC showed that the ratio immediately after the reaction was 1:1, indicating that the ratio of 1:0.7 is not the result of a kinetically controlled reaction, but the result of equilibrium after thermodynamic conversion.

Both isomers were subjected to X-ray structure analysis to reveal that the benzolactam moiety exists in a boat-like form, and the absolute stereochemistry at the axis is aR for 7a-A and aS for 7a-B (Figure 5).^{14,15} As shown in Figure 5, the isomers adopt antipode forms at the benzolactam moiety, and the phenyl group of the phenethylamide moiety of 7a-A locates anti to the benzene ring of the benzolactam moiety (extended form), whereas that of 7a-B adopts the syn (folded form). The extended form seemed to be thermodynamically more stable than the folded form to result in the preferred formation of 7a-A. In the ¹H NMR spectra of **7a-A** and **7a-B** (Figure 6), the same coupling patterns of the ring methylene protons were observed, which indicates that both diastereomers have the same ring conformation (antipode form) in solution (CDCl₃). As shown in Figure 6, the ¹H NMR spectra are diagnostic for determining the stereochemistry. Thus, the methylene protons of 7a-A appear in a higher field compared



Figure 6. ¹H NMR spectra of 7a-A (aR, S) (above) and 7a-B (aS, S) (below).



Figure 7. X-ray crystal structure of 7b-A (aR, S).

with those of 7**a**-**B**: H^{3a} , H^{3b} , H^{4a} , H^{4b} : δ (ppm) 2.34, 2.43, 2.70, 3.28 for 7**a**-**A**, and 2.39, 2.51, 2.94, 3.65 for 7**a**-**B**, respectively. These data may easily be explained as due to the methylene protons of the lactam ring of 7**a**-**A** locating over the benzene ring in the phenethylamide moiety to appear at a higher field.

Similarly, compound **8a** (Y = Ph) was obtained as a mixture of the diastereomers in a ratio of A:B = 1:0.9, which were separated by column chromatography (SiO₂). The stereochemistry was assigned on the basis of the chemical shifts of the methylene protons as described above.

Compound 7b (X = S, Y = Cl) was obtained as a mixture of the diastereomers in a ratio of 1:0.9, which were separated by crystallization to afford 7b-A as crystals (yield 37%). From the mother liquor, the isomer 7b-B (in 85% de) was obtained as an oily substance. The stereochemistry was clarified by X-ray structure analysis of 7b-A (Figure 7), which showed that the thiazepinone ring takes a boatlike form similar to 7a; the stereochemistry at the axis of 7b-A is aR, ¹⁵ and hence, that of 7b-B was determined to be a*S*. The same tendency of the chemical shifts of the methylene protons in the ¹H NMR spectra of 7b-A and B as those in 7a-A and B were observed, e.g., H^{2a}: δ (ppm) 3.15 for 7b-A, and 3.27 for 7b-B.

Compound 7c (X = CH₂) was also obtained as a mixture of diastereomers in a ratio of 1:0.8, which were separated by column chromatography (SiO₂) to afford 7c-A (yield 36%) and 7c-B (yield 31%) as crystals. The stereochemistry was deduced by comparing the chemical shifts of the methylene protons in the ¹H NMR spectra as discussed above, e.g., H^{3a}, H^{3b}: δ (ppm) 2.37, 2.84 for 7c-A, and 2.65, 3.44 for 7c-B.

Stereochemical Stability of Atropisomers of Seven-Membered-Ring Benzolactams (7a-c and 8a). The stereochemical

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Table 1. Separation of the Atropisomers (A and B) and Their Stereochemical Stability



compd (axial chirality ^a)	mp (°C)	ΔG^{\ddagger} kJ/mol	conditions for isomerization b	equilibrium ratio (A:B)
7 a-A (aR)	173-175	102.5	37 °C, 10 h ^c	1:0.7
B (aS)	124-127	99.4	37 °C, 8 h	
8a-A (aS)	196-198	106.9	50 °C, 14 h ^d	1:0.9
B (a <i>R</i>)	196-198	105.8	50 °C, 12 h ^e	
7 b-A (a <i>R</i>)	142-144	100.8	37 °C, 8 h	1:0.9
B (aS)	oil ^f	99.2	37 °C, 7 h	
7 c-A (a <i>R</i>)	52-54	101.9	37 °C, 9 h	1:0.8
B (aS)	78-80	99.8	37 °C, 8 h	
^{<i>a</i>} See ref 14. ^{<i>b</i>} To the equilibriu ^{<i>f</i>} 85% de.	um state in toluene. ^{<i>c</i>} At	50 °C, 2 h. ^{<i>d</i>} At 37 °C afte	er 7 h, isomerized to 71% de. ^{<i>e</i>} At 37 °C at	fter 7 h, isomerized to 60% de.

stability of these separated diastereomers of $7\mathbf{a} - \mathbf{c}$ and $8\mathbf{a}$ was next examined at 37 °C (or 50 °C) in toluene. The activation free energy barrier to rotation $(\Delta G^{\ddagger})^{17}$ and the conditions required for isomerization to the equilibrium state are shown in Table 1. The diastereomers $7\mathbf{a}$ -**A** and **B** showed ΔG^{\ddagger} values of 102.5 and 99.4 kJ/ mol, respectively, which may reflect the equilibrium ratio of 1:0.7. The higher stereochemical stability of $8\mathbf{a}$ -**A** (106.9 kJ/mol) compared with $7\mathbf{a}$ -**A** (102.5 kJ/mol) is well explained because of the steric bulkiness of phenyl vs Cl. The stability of benzolactams $7\mathbf{b}$ (**A**/**B**) and $7\mathbf{c}$ (**A**/**B**) was revealed to be almost the same as that of $7\mathbf{a}$ (**A**/**B**), which reached the equilibrium state at 37 °C for 7 to 10 h. Here again, the energy barrier to rotation of type **A** was shown to be slightly higher than that of type **B**.

CONCLUSION

In conclusion, atropisomers of seven-membered-ring benzolactams (I) (1,5-benzodiazepin-2-one, 1,5-benzothiazepin-4-one, and 1-benzazepin-2-one) were separated as diastereomers, and the conformation was first clarified from the viewpoint of the atropisomerism. The axial chirality at the aryl—amide bond is often overlooked, although it exists in various forms as observed in the dibenzo[b,d]azepin-6-ones (II). These results may support our assumption that the two axial chiralities exist in II and move together like a gear and may prove the relevance of the atropisomeric structures of these heterocycles to the biological activity. We hope that this work provides useful information for future drug design.

EXPERIMENTAL SECTION

The NH-lactams 5-methyl-1,3,4,5-tetrahydro-2*H*-1,5-benzodiazepin-2-one (1a),¹⁰ 2,3-dihydro-1,5-benzo[b][1,4]thiazepine-4(5*H*)-one (1b),¹¹

and 1,3,4,5-tetrahydro-2*H*-1-benzoazepin-2-one $(1c)^{12}$ were prepared according to the reported procedure, which were used for the preparation of the benzolactams with a chloro substituent at the ortho position of the benzene ring (2a, 2b, and 2c) as described below.

9-Chloro-5-methyl-1,3,4,5-tetrahydro-2*H***-1,5-benzodiazepin-2-one (2a).** (1) Concentrated H₂SO₄ (98%, 15 mL) was added by portions to a mixture of 3-chloro-*N*-methyl-2-nitroaniline¹⁸ (5.75 g, 30.8 mmol) and acrylic acid (25 mL, 308 mmol) in H₂O (27 mL). The mixture was stirred at 80 °C for 5 h. After being cooled to room temperature, the mixture was extracted with EtOAc. The extract was washed with H₂O and brine and dried. Removal of the solvent gave yellow crystals which were collected and washed with *i*-Pr₂O to afford *N*-(3-chloro-2-nitrophenyl)-*N*-methyl-*β*-alanine (5.59 g, 70%): mp 126–129 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.55 (t, *J* = 7.3 Hz, 2H), 2.77 (s, 3H), 3.34 (t, *J* = 7.3 Hz, 2H), 7.16 (dd, *J* = 0.9, 8.0 Hz, 1H), 7.17 (dd, *J* = 0.9, 8.3 Hz, 1H), 7.34 (dd, *J* = 8.0, 8.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 32.6, 42.3, 51.4, 121.1, 124.6, 125.8, 130.8, 145.6, 176.6; IR (KBr) 2966, 1714, 1594 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₀H₁₀N₂O₄Cl 257.0335 (M – H)⁻, found 257.0349.

(2) To a stirred solution of *N*-(3-chloro-2-nitrophenyl)-*N*-methyl- β -alanine (250 mg, 0.97 mmol) in 1,4-dioxane (2.4 mL) and H₃PO₄ (85%, 0.57 mL) was added Zn powder (474 mg, 7.25 mmol) by portions at 80 °C. The mixture was refluxed for 5 h. After being cooled to room temperature, the mixture was filtered. The filtrate was poured into aq NaHCO₃, and the aqueous mixture was extracted with EtOAc. The extract was washed with H₂O and brine and dried. Removal of the solvent gave colorless crystals that were collected and washed with *i*-Pr₂O to afford **2a** (114 mg, 56%): mp 159–162 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.52 (t, *J* = 7.3 Hz, 2H), 2.84 (s, 3H), 3.54 (t, *J* = 7.3 Hz, 2H), 6.95 (dd, *J* = 1.7, 7.3 Hz, 1H), 7.07 (dd, *J* = 1.7, 8.0 Hz, 1H), 7.09 (dd, *J* = 7.3, 8.0 Hz, 1H), 7.22 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.7, 41.5, 58.2, 118.1, 122.7, 125.9, 126.1, 129.4, 144.0, 172.4; IR (KBr) 3178, 1680 cm⁻¹; HRMS (ESI) m/z calcd for C₁₀H₁₂N₂OCl 211.0633 (M + H)⁺, found 211.0632.

5-Methyl-9-phenyl-1,3,4,5-tetrahydro-2H-1,5-benzodiazepin-2-one (3a). To a stirred solution of 2a (594 mg, 2.82 mmol) in a mixture of N,N-dimethylacetamide (DMA) (17.1 mL) and H₂O (0.9 mL) were added tris(dibenzylideneacetone)dipalladium (0) $(Pd_2(dba)_3)$ (65 mg, 2.5 mol %), 2-dicyclohexcylphosphino-2',4',6'triisopropylbiphenyl (Xphos) (81 mg, 6 mol %), and K₂CO₃ (779 mg, 5.64 mmol). After being stirred at 25 °C for 1 h under argon, the mixture was treated with phenylboronic acid (1.03 g, 8.46 mmol) and stirred at 100 °C for 7 h under argon. After being cooled to room temperature, the mixture was filtered through Celite. The filtrate was poured into H₂O, and the aqueous mixture was extracted with EtOAc. The extract was washed with brine and dried. After removal of the solvent, the residue was purified by column chromatography (silica gel, EtOAc/hexane = 1/3) to afford 3a as colorless crystals (656 mg, 92%): mp 127-130 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.60 (t, J = 7.0 Hz, 2H), 2.88 (s, 3H), 3.56 (t, J = 7.0 Hz, 2H), 6.71 (br,1H), 6.98 (dd, J = 1.4, 7.5 Hz, 1H), 7.07 (dd, *J* = 1.2, 8.0 Hz, 1H), 7.24 (dd, *J* = 7.3, 7.8 Hz, 1H), 7.30–7.32 (m, 2H), 7.35–7.39 (m, 1H), 7.41–7.45 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 33.9, 41.6, 58.5, 118.9, 123.8, 126.0, 127.8, 128.8, 129.2, 134.7, 137.9, 143.0, 172.9; IR (KBr) 3055, 1670 cm⁻¹; HRMS (ESI) *m/z* calcd for $C_{16}H_{17}N_2O$ 253.1335 (M + H)⁺, found 253.1333.

6-Chloro-2,3-dihydro-1,5-benzo[b][1,4]thiazepine-4(5H)one (2b). To a stirred solution of 2-amino-3-chlorothiophenol¹⁹ (0.80 g, 5.0 mmol) in pyridine (6.0 mL) was added β -propiolactone (0.31 mL, 5.0 mmol) at 25 °C, and the mixture was stirred for 30 min. To the mixture was added acetic anhydride (6.0 mL) dropwise over a period of 1 h with stirring. After being stirred at 5 °C for 14 h, the mixture was concentrated. To the concentrate was added EtOAc, and the mixture was washed successively with H2O, 2 N HCl, H2O, aq NaHCO3, and H2O and dried. The organic layer was evaporated to afford 2b as colorless crystals (755 mg, 3.53 mmol, 71%): mp 168-170 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.64 (t, J = 6.9 Hz, 2H), 3.45 (t, J = 6.9 Hz, 2H), 7.10 (t, J = 7.9 Hz, 1H), 7.30(s, 1H), 7.42 (dd, J = 1.4, 7.9 Hz, 1H), 7.51 (dd, J = 1.4, 7.9 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.7, 34.5, 126.6, 127.2, 128.6, 130.3, 134.1, 138.8, 172.2; IR (KBr) 3112, 1672 cm⁻¹ HRMS (ESI) m/z calcd for C₉H₈ClNOS 214.0080 (M + H)⁺, found 214.0088.

2,3-Dihydro-6-phenyl-1,5-benzo[b][1,4]thiazepine-4(5H)one (3b). To a stirred solution of 2b (106 mg, 0.5 mmol) in a mixture of DMA (3.0 mL) and H₂O (0.16 mL) were added Pd₂(dba) (46 mg, 10 mol %), Xphos (57 mg, 24 mol %), and K₂CO₃ (138 mg, 1.0 mmol). After being stirred at room temperature for 1 h under argon atmosphere, the mixture was treated with phenylboronic acid (487 mg, 4.0 mmol) and stirred at 100 °C for 7 h under argon atmosphere. After being cooled to room temperature, the mixture was filtered through Celite. The filtrate was poured into H2O. The aqueous mixture was extracted with EtOAc, and the extract was washed with H₂O and dried. After removal of the solvent, the oily residue was purified by column chromatography (silica gel, EtOAc/hexane = 1:3) to give 3b as colorless crystals (123 mg, 97%): mp 175–177 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.74 (t, J = 6.8 Hz, 2H), 3.48 (t, J = 6.8 Hz, 2H), 6.86 (br, 1H), 7.23 (t, J = 7.5 Hz, 1H), 7.28–7.34 (m, 3H), 7.36–7.47 (m, 3H), 7.63 (dd, J = 1.4, 7.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.7, 34.3, 100.5, 126.2, 127.2, 128.1, 128.9, 129.0, 131.1, 134.7, 135.9, 138.6, 172.4; IR (KBr) 2918, 1674 cm $^{-1};$ HRMS (ESI) $\mathit{m/z}$ calcd for $\rm C_{15}H_{13}NOS$ $256.0791 (M + H)^+$, found 256.0793.

9-Chloro-1,3,4,5-tetrahydro-2*H*-1-benzoazepin-2-one (2c) and 2c-Regioisomer via the Beckmann Rearrangement of 8-Chloro-3,4-dihydronaphthalen-1(2*H*)-one Oxime. (1) A mixture of 8-chloro-3,4-dihydro-1(2*H*)-naphthalenone²⁰ (6.30 g, 34.9 mmol), hydroxylamine hydrochloride (7.20 g, 104 mmol), sodium acetate (8.64 g, 105 mmol), and EtOH (350 mL) was refluxed for 1.5 h.

After being cooled to room temperature, the mixture was poured into ice—water. Pale yellow crystals separated were collected by filtration and washed with H₂O and hexane to afford 8-chloro-3,4-dihydronaphthalen-1(2*H*)-one oxime (6.30 g, 92%): mp 159–161 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.79 (tt, *J* = 6.1, 6.8 Hz, 2H), 2.65 (t, *J* = 6.1 Hz, 2H), 2.87 (t, *J* = 6.8 Hz, 2H), 7.07 (dd, *J* = 0.7, 7.5 Hz, 1H), 7.14 (dd, *J* = 7.5, 8.0 Hz, 1H), 7.32 (dd, *J* = 0.7, 8.0 Hz, 1H), 9.30 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.0, 24.8, 31.0, 126.4, 128.8, 128.9, 129.5, 131.4, 143.8, 153.9; IR (KBr) 3231 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₀H₁₀ClNO 196.0524 (M + H)⁺, found 196.0524. The oxime thus obtained is the sole isomer, whose stereochemistry is presumed to be (*E*).

(2) The mixture of the oxime (6.00 g, 30.6 mmol) and polyphosphoric acid (PPA) (104 g) was stirred at 95 °C for 1 h and then cooled to room temperature. The mixture was poured into ice-water and extracted with EtOAc. The extract was washed successively with H₂O, saturated NaHCO3, and H2O and dried. Removal of the solvent gave a white solid, which was purified by column chromatography (silica gel, EtOAc/hexane = 1/4 to 1/1) to afford 2c (3.10 g, 52%) and 9-chloro-2,3,4,5-tetrahydro-1H-2-benzoazepin-1-one (2c-regio isomer) (2.80 g, 47%) as colorless crystals. Data for 2c: mp 162-164 °C; ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3) \delta 2.26 (\text{quin}, J = 6.8, 7.2 \text{ Hz}, 2\text{H}), 2.38 (t, J = 7.2 \text{ Hz}, 2\text{Hz})$ 2H), 2.83 (t, J = 6.8 Hz, 2H), 7.08 (t, J = 7.8 Hz, 1H), 7.14 (dd, J = 1.4, 7.8 Hz, 1H), 7.22 (1H, b), 7.31 (dd, J = 1.4, 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 28.4, 30.8, 32.8, 125.8, 126.0, 127.8, 128.2, 134.8, 136.0, 173.7; IR (KBr) 3190, 1656 cm⁻¹; HRMS (ESI) m/z calcd for $C_{10}H_{10}CINO$ 196.0524 (M + H)⁺, found 196.0524. Data for 2cregioisomer: mp 192–195 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.96 (br, 2H), 2.83 (br, 2H), 3.11 (br, 2H), 6.51 (br, 1H), 7.10 (dd, J = 0.8, 7.8 Hz, 1H), 7.29 (t, J = 7.8 Hz, 1H), 7.37 (dd, J = 0.8, 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 30.1, 30.3, 39.0, 126.7, 128.9, 131.0, 132.4, 132.9, 139.4, 169.5; IR (KBr) 3190, 1656 cm $^{-1}$; HRMS (ESI) m/z calcd for $C_{10}H_{10}CINO$ 196.0524 $(M + H)^+$, found 196.0526.

9-Phenyl-1,3,4,5-tetrahydro-2H-1-benzoazepin-2-one (3c). To a stirred solution of 2c (81 mg, 0.41 mmol) in a mixture of DMA (2.8 mL) and H₂O (0.25 mL) were added Pd₂(dba)₃ (57 mg, 0.060 mmol), Xphos (74 mg, 0.015 mmol), and K₂CO₃ (114 mg, 0.830 mmol). After being stirred at 25 °C for 1 h under argon atmosphere, the mixture was treated with phenylboronic acid (91 mg, 0.75 mmol) and stirred at 100 °C for 50 h under argon atmosphere. After being cooled to room temperature, the mixture was filtered through Celite, and the filtrate was poured into H₂O. The aqueous mixture was extracted with EtOAc, and the extract was washed with brine, dried, and evaporated. After removal of the solvent, the residue was purified by column chromatography (silica gel, EtOAc/hexane = 1/6) to afford 3c as a white solid (87 mg, 88%): mp 148–149 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.29 (tt, J = 7.0, 7.3 Hz, 2H), 2.45 (t, J = 7.3 Hz, 2H), 2.84 (t, J = 7.0 Hz, 2H), 6.83 (br, 1H), 7.20-7.25 (m, 3H), 7.29-7.31 (m, 2H), 7.35-7.39 (m, 1H), 7.41-7.46 (m, 2H); $^{13}{\rm C}$ NMR (100 MHz, CDCl_3) δ 28.4, 30.6, 32.8, 125.5, 127.6, 128.6, 128.7, 128.9, 134.3, 134.6, 134.7, 137.7, 174.0; IR (KBr) 3054, 1664 cm^{-1} ; HRMS (ESI) m/z calcd for $C_{16}H_{15}NO 238.1226 (M + H)^+$, found 238.1237.

Preparation of Methyl (1,5-Benzodiazepin-1-yl)acetates 4a, 5a, and 6a: Methyl (5-methyl-2-oxo-2,3,4,5-tetrahydro-1*H*-1,5-benzodiazepin-1-yl)acetate (4a). To a solution of 1a¹⁰ (1.00 g, 5.67 mmol) in DMF (16 mL) was added NaH (60%, 681 mg, 17 mmol) at -20 °C, and the mixture was stirred at -20 °C for 15 min. Then methyl bromoacetate (2.7 mL, 28.4 mmol) was added to the mixture, and stirring was continued at -20 °C for 2.5 h. After being warmed to room temperature, the mixture was poured into H₂O. The aqueous mixture was extracted with EtOAc, and the combined organic layer was washed with H₂O and dried. After removal of the solvent, the residue was purified by column chromatography (silica gel, EtOAc/ dichloromethane =1:5) to give 4a as a colorless oil (1.26 g, 89%): ¹H NMR (400 MHz, CDCl₃) δ 2.52 (t-br, J = 6.8 Hz, 2H), 2.81 (s, 3H), 3.43 (br, 2H), 3.80 (s, 3H), 4.41 (br, 2H), 7.01–7.06 (m, 2H), 7.14 (dd, J = 1.4, 7.8 Hz, 1H), 7.22 (ddd, J = 1.4, 7.8, 8.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 34.0, 41.0, 50.9, 52.4, 58.7, 119.3, 122.3, 122.8, 137.5, 143.2, 170.0, 172.3; IR (neat) 3489, 1752, 1676 cm⁻¹; HRMS (ESI) m/z calcd for C₁₃H₁₇N₂O₃ 249.1234 (M + H)⁺, found 249.1235.

Compounds 5a and 6a were prepared from the corresponding 1,5benzodiazepinones (2a, 3a) according to a similar procedure described for the preparation of 4a from 1a.

Methyl (9-chloro-5-methyl-2-oxo-2,3,4,5-tetrahydro-1*H*-1,5-benzodiazepin-1-yl)acetate (5a): colorless oil (97%); ¹H NMR (400 MHz, CDCl₃) δ 2.39 (ddd, *J* = 1.2, 5.6, 12.9 Hz, 1H), 2.57 (ddd, *J* = 7.0, 12.9, 13.1 Hz, 1H), 2.84 (s, 3H), 3.09 (ddd, *J* = 1.2, 7.0, 10.0 Hz, 1H), 3.70 (s, 3H), 3.74 (ddd, *J* = 5.6, 10.0, 13.1 Hz, 1H,), 4.24 (d, *J* = 17.0 Hz, 1H), 4.53 (d, *J* = 17.0 Hz, 1H), 7.00 (dd, *J* = 1.4, 8.0 Hz, 1H), 7.12 (dd, *J* = 1.4, 8.0 Hz, 1H), 7.19 (dd, *J* = 8.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.6, 41.1, 49.6, 52.0, 58.2, 118.2, 124.4, 128.2, 130.4, 133.9, 146.6, 168.6, 173.1; IR (neat) 3355, 1750, 1681 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₃H₁₆N₂O₃Cl 283.0844 (M + H)⁺, found 283.0830.

Methyl (5-methyl-2-oxo-9-phenyl-2,3,4,5-tetrahydro-1*H*-1,5-benzodiazepin-1-yl)acetate (6a): colorless crystals (82%); mp 157–159 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.49 (dd, *J* = 5.3, 12.7 Hz, 1H), 2.84 (ddd, *J* = 7.3, 12.7, 13.1 Hz, 1H), 2.90 (s, 3H), 3.18 (d, *J* = 17.0 Hz, 1H), 3.20 (dd, *J* = 7.3, 10.0 Hz, 1H), 3.52 (s, 3H), 3.81 (ddd, *J* = 5.3, 10.0, 13.1 Hz, 1H), 4.15 (d, *J* = 17.0 Hz, 1H), 7.04 (dd, *J* = 1.4, 7.5 Hz, 1H), 7.09 (dd, *J* = 0.9, 8.3 Hz, 1H), 7.31–7.37 (m, 4H), 7.40–7.43 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 34.1, 41.1, 49.4, 51.7, 58.2, 118.7, 125.0, 127.7, 127.9, 128.4, 128.9, 134.0, 136.6, 139.0, 145.4, 168.6, 173.5; IR (KBr) 2949, 1737, 1669 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₉H₂₁N₂O₃ 325.1547 (M + H)⁺, found 325.1531.

Preparation of Methyl (1,5-Benzothiazepinyl)acetate: 4b, 5b, and 6b. Compounds **4b**, **5b**, and **6b** were prepared from the corresponding 1,5-benzothiazepinones (1b,¹¹ 2b, 3b) according to a similar procedure described for the preparation of **4a** from **1a**.

Methyl (3,4-dihydro-4-oxo-1,5-benzothiazepin-5(2*H*)yl)acetate (4b): colorless oil (97%); ¹H NMR (600 MHz, CDCl₃) δ 2.65 (br, 2H), 3.40 (br, 2H), 3.80 (s, 3H), 4.00 (br, 1H), 4.90 (br, 1H), 7.22 (ddd, *J* = 1.3, 7.4, 7.4 Hz, 1H), 7.33 (dd, *J* = 1.3, 7.4 Hz, 1H), 7.41 (ddd, *J* = 1.6, 7.4, 7.4 Hz, 1H), 7.62 (dd, *J* = 1.6, 7.4 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.8, 34.1, 51.0, 52.4, 123.6, 127.2, 127.7, 130.7, 135.7, 146.8, 169.7, 171.7; IR (neat) 2953, 1750, 1672 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₂H₁₃NO₃S 252. 0685 (M + H)⁺, found 252.0694.

Methyl (6-chloro-3,4-dihydro-4-oxo-1,5-benzothiazepin-5(*2H*)-yl)acetate (5b): colorless crystals (98%); mp 93–96 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.54 (ddd, *J* = 7.6, 12.4, 12.6 Hz, 1H), 2.67 (ddd, *J* = 1.8, 6.0, 12.6 Hz, 1H), 3.29–3.41 (m, 2H), 3.73 (s, 3H), 4.29 (d, *J* = 16.8 Hz, 1H), 4.54 (d, *J* = 16.8 Hz, 1H), 7.22 (dd, *J* = 7.5, 8.0 Hz, 1H), 7.49 (dd, *J* = 1.2, 8.0 Hz, 1H), 7.59 (dd, *J* = 1.2, 7.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.4, 33.9, 49.4, 52.0, 128.5, 129.9, 131.8, 132.1, 134.7, 168.2, 172.1; IR (KBr) 2951, 1761, 1735 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₂H₁₂NO₃SCl 308.0119 (M + Na)⁺, found 308.0107. This compound was subjected to X-ray crystallographic analysis.

Methyl (3,4-dihydro-4-oxo-6-phenyl-1,5-benzothiazepin-5(*2H*)-yl)acetate (6b): white solid (76%); mp 136–138 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.76 (ddd, *J* = 1.9, 6.3, 12.4 Hz, 1H), 2.84 (ddd, *J* = 7.0, 12.4, 12.4 Hz, 1H), 3.23 (d, *J* = 16.8 Hz, 1H), 3.39 (ddd, *J* = 6.3, 12.4, 12.4 Hz, 1H), 3.47 (ddd, *J* = 1.9, 7.0, 12.4 Hz, 1H), 3.55 (s, 3H), 4.16 (d, *J* = 16.8 Hz, 1H), 7.29–7.46 (m, 7H), 7.66 (dd, *J* = 1.7, 7.5 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.7, 33.8, 49.5, 51.7, 127.9, 128.1, 128.1, 129.1, 130.7, 132.3, 135.1, 137.5, 138.3, 168.3, 172.5; IR (KBr) 2957, 1737, 1670 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₈H₁₇NO₃S 328.1002 (M + H)⁺, found 328.1000. Preparation of Methyl (1-Benzoazepinyl)acetate: 4c, 5c, and 6c. Compounds 4c, 5c, and 6c were prepared from the corresponding 1-benzoazepinones $(1c, ^{12} 2c, 3c)$ according to a similar procedure described for the preparation of 4a from 1a.

Methyl (2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzoazepin-1-yl)acetate (4c): white solid (93%); mp 79–80 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.20 (br, 2H), 2.35 (br-t, *J* = 6.8 Hz, 2H), 2.92 (br, 2H), 3.74 (s, 3H), 4.51 (s, 2H), 7.11–7.22 (m, 3H), 7.25–7.29 (m, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 28.8, 29.8, 32.8, 49.8, 52.2, 122.0, 126.4, 127.4, 129.4, 135.8, 142.2, 169.6, 173.2; IR (KBr) 2954, 1746, 1672 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₃H₁₅NO₃ 234.1125 (M + H)⁺, found 234.1125.

Methyl (9-chloro-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzoazepin-1-yl)acetate (5c): white solid (90%); mp 92–94 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.88–1.98 (m, 1H), 2.22–2.44 (m, 3H), 2.65 (dd, *J* = 6.5, 13.4 Hz, 1H), 3.48–3.72 (m, 1H), 3.69 (s, 3H), 4.11 (d, *J* = 17.3 Hz, 1H), 4.88 (d, *J* = 17.3 Hz, 1H), 7.14–7.20 (m, 2H), 7.34 (dd, *J* = 2.1, 7.3 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 28.3, 29.7, 32.2, 49.3, 52.0, 127.9, 128.3, 128.7, 129.0, 138.7, 140.0, 169.5, 173.5; IR (KBr) 2950, 1742, 1673 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₃H₁₄NO₃Cl 290.0554 (M + Na)⁺, found 290.0554.

Methyl (2-oxo-9-phenyl-2,3,4,5-tetrahydro-1*H*-1-benzoazepin-1-yl) acetate (6c): hite solid (97%); mp 150–153 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.99–2.09 (m, 1H), 2.41–2.49 (m, 2H), 2.53–2.59 (m, 1H), 2.69 (dd, *J* = 6.3, 13.4 Hz, 1H), 3.11 (d, *J* = 17.3 Hz, 1H), 3.59 (s, 3H), 3.70 (ddd, *J* = 6.8, 7.0, 13.4 Hz, 1H), 4.17 (d, *J* = 17.3 Hz, 1H), 7.23 (dd, *J* = 2.6, 6.5 Hz, 1H), 7.27–7.32 (m, 4H), 7.34–7.38 (m, 1H), 7.41–7.45 (m, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 28.5, 29.7, 32.7, 49.6, 51.8, 127.5, 127.7, 128.2, 128.7, 129.0, 129.7, 136.0, 138.1, 138.9, 139.0, 169.4, 174.0; IR (KBr) 2952, 1755, 1666 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₉H₁₉NO₃ 332.1257 (M + Na)⁺, found 332.1272.

2-(9-Chloro-5-methyl-2-oxo-2,3,4,5-tetrahydro-1H-1,5benzodiazepin-1-yl)-*N*-[(1*S*)-1-phenylethyl]acetamide (7a) and the Atropisomers 7a-A and 7a-B. 1) To a solution of 5a (1.00 g, 3.54 mmol) in MeOH (10 mL) was added 1 N NaOH aq (30 mL). After being stirred at 25 °C for 2 h, the mixture was acidified with 2 N HCl and extracted with EtOAc. The extract was dried and concentrated to afford 9-chloro-5-methyl-2-oxo-2,3,4,5-tetrahydro-1H-1,5-benzodiazepin-1-yl)acetic acid (5a-CO₂H) as a white solid (662 mg, 70%): mp 145–147 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.47 (ddd, J = 0.9, 6.1, 13.1 Hz, 1H), 2.54 (ddd, J = 7.3, 12.4, 13.1 Hz, 1H), 2.89 (s, 3H), 3.06 (ddd, J = 0.9, 7.3, 10.5 Hz, 1H), 3.78 (ddd, J = 6.1, 10.5, 12.4 Hz, 1H), 4.01 (d, J = 17.0 Hz, 1H), 5.06 (d, J = 17.0 Hz, 1H), 7.20 (dd, J = 2.6, 6.5 Hz, 1H, 7.37 - 7.42 (m, 2H), 12.43 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 32.9, 40.6, 50.1, 58.9, 118.5, 127.6, 129.8, 130.4, 134.4, 144.6, 169.1, 172.2; IR (KBr) 3334, 1750, 1683 cm⁻¹; HRMS (ESI) m/zcalcd for 269.0687 $C_{12}H_{14}N_2O_3Cl (M + H)^+$, found 269.0679.

(2) To a solution of 5a-CO₂H (150 mg, 0.558 mmol) in THF (4.5 mL) were added N-hydroxybenzotriazole (HOBt) (82.9 mg, 0.614 mmol), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDC) (321 mg, 1.67 mmol), and (S)-(-)-1-phenethylamine ((S)-PEA) (0.36 mL, 2.8 mmol). The mixture was stirred at 25 °C for 3 h. After removal of the solvent, the residue was poured into aq NaHCO3. The aqueous solution was extracted with EtOAc, and the extract was washed successively with aq NaHCO₃, H₂O, dilute HCl, and H₂O and dried. Removal of the solvent gave a mixture of the diastereomers 7a-A and 7a-B (ratio 1: 0.67 determined by ¹H NMR) as a white foam (166 mg, 80%). The mixture was separated and purified by column chromatography (silica gel, EtOAc/ hexane = 1:2) to give 7a-A (aR,S) (isolated yield: 69 mg, 34%) and 7a-B (aS,S) (isolated yield: 45 mg, 22%). 7a-A (aR,S): colorless crystals; mp 173–175 °C; $[\alpha]^{21}_{D}$ +111.2 (*c* 0.30, CHCl₃); ¹H NMR (400 MHz, $CDCl_3$) δ 1.37 (d, J = 6.8 Hz, 3H), 2.19 (s, 3H), 2.34 (dd, J = 5.9, 12.9 Hz, 1H), 2.43 (ddd, J = 5.9, 7.3, 12.9 Hz, 1H), 2.70 (dd, J = 7.3, 10.3 Hz, 1H), 3.28 (ddd, *J* = 5.9, 10.3, 12.9 Hz, 1H), 4.02 (d, *J* = 17.3 Hz, 1H), 4.91 (d, J = 17.3 Hz, 1H), 5.17 (quin, J = 6.8 Hz, 1H), 6.99 (t, J = 4.6 Hz, 1H), 7.26-7.27 (m, 3H), 7.38-7.40 (m, 4H), 8.30 (br-d, J = 6.8 Hz, 1H); 13 C NMR (100 MHz, CDCl₃) δ 20.2, 33.2, 40.2, 47.1, 51.9, 58.5, 118.1, 126.5, 127.2, 128.4, 128.9, 129.9, 134.3, 142.6, 145.4, 167.3, 171.8; IR (KBr) 3236, 1685, 1664 cm⁻¹; HRMS (ESI) m/z calcd for $C_{20}H_{23}N_3O_2Cl$ 372.1473 (M + H)⁺, found 372.1454. 7a-B (aS,S): colorless crystals; mp 124–127 °C; $[\alpha]^{21}_{D}$ –44.9 (*c* 0.30, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.51 (d, J = 7.1 Hz, 3H), 2.25 (s, 3H), 2.39 (dd, J = 5.6, 12.9 Hz, 1H), 2.51 (ddd, J = 5.67.3, 12.9 Hz, 1H), 2.94 (dd, J = 7.3, 10.5 Hz, 1H), 3.65 (ddd, J = 5.6, 10.5, 12.9 Hz, 1H), 4.08 (d, J = 17.3 Hz, 1H), 4.98 (d, J = 17.3 Hz, 1H), 5.15 (quin, J = 7.1 Hz, 1H), 6.92 (dd, J = 4.6, 8.8 Hz, 1H), 7.03-7.06 (m, 2H), 7.17-7.22 (m, 2H),7.26–7.27 (m, 3H), 8.08 (br-d, J = 7.1 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.0, 33.3, 40.3, 47.5, 51.2, 59.2, 118.0, 126.2, 126.5, 127.0, 128.3, 128.8, 129.8, 133.9, 143.0, 145.4, 167.2, 171.8; IR (KBr) 3246, 1682, 1670 cm⁻¹; HRMS (ESI) m/z calcd for C₂₀H₂₃N₃O₂Cl 372.1473 $(M + H)^+$, found 372.1454.

2-(6-Chloro-3,4-dihydro-4-oxo-1*H*-1,5-benzothiazepin-1yl)-*N*-[(15)-1-phenylethyl]acetamide (7b) and the Atropisomers 7b-A and 7b-B. Compounds 7b was prepared from the corresponding acetate (5b) via the carboxylic acid (5b-CO₂H) according to a similar procedure described for the preparation of 7a from Sa.

(6-Chloro-3,4-dihydro-4-oxo-1*H*-1,5-benzothiazepin-1-yl)acetic acid (5b-CO₂H): white solid (88%); mp 144–146 °C; ¹H NMR (600 MHz, DMSO- d_6) δ 2.41 (ddd, J = 7.3, 12.8, 12.8 Hz, 1H), 2.51 (ddd, J = 1.1, 6.2, 12.8 Hz, 1H), 3.21 (ddd, J = 6.2, 11.7, 12.8 Hz, 1H), 3.33 (ddd, J = 1.1, 7.3, 11.7 Hz, 1H), 4.13 (d, J = 17.2 Hz, 1H), 4.17 (d, J = 17.2 Hz, 1H), 7.34 (dd, J = 7.6, 8.4 Hz, 1H), 7.61 (dd, J = 1.4, 7.6 Hz, 1H), 7.64 (dd, J = 1.4, 8.4 Hz, 1H), 12.4 (br, 1H); ¹³C NMR (150 MHz, *d*-DMSO) δ 32.9, 33.5, 49.3, 128.9, 129.1, 131.2, 132.0, 134.7, 142.9, 168.5, 171.6; IR (KBr) 2918, 1747, 1632 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₁H₁₀NO₃SCI 293.9962 (M + Na)⁺, found 293.9947.

2-(6-Chloro-3,4-dihydro-4-oxo-1H-1,5-benzothiazepin-1yl)-N-[(1S)-1-phenylethyl]acetamide (7b). The mixture (with a ratio of 1:0.9 determined by ¹H NMR) of the diastereomers of 7b-A and 7b-B was obtained as a colorless foam (yield 99%). The foam was treated with Et_2O to separate the diastereomers 7b-A (aR,S) and 7b-B (aS,S). The diastereomer 7b-A was obtained as colorless crystals (isolated yield: 37%), and the diastereomer 7b-B was obtained from the mother liquor as a colorless oily substance (isolated yield: 40% in a 85% de (B:A = 5.6:1)). 7**b-A** (a*R*,*S*): colorless crystals; mp 142–144 °C; $[\alpha]_{D}^{25}$ –107.8 (*c* 0.1, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 1.45 (d, *J* = 7.2 Hz, 3H), 2.46 (ddd, *J* = 7.2, 12.8, 12.8 Hz, 1H), 2.64 (ddd, *J* = 1.2, 5.6, 12.8 Hz, 1H), 3.15 (ddd, J = 5.6, 12.8, 12.8 Hz, 1H), 3.35 (ddd, J = 1.2, 7.2, 12.8 Hz, 1H), 4.07 (d, J = 16.6 Hz, 1H), 4.84 (d, J = 16.6 Hz, 1H), 5.11 (quin, J = 7.2 Hz, 1H), 7.25-7.30 (m, 2H), 7.33-7.36 (m, 4H), 7.55 (dd, *J* = 1.2, 8.2 Hz, 1H), 7.59 (dd, *J* = 1.2, 7.6 Hz, 1H), 7.90 (br-d, *J* = 7.6 Hz, 1H); 13 C NMR (100 MHz, CDCl₃) δ 21.7, 33.1, 34.1, 48.9, 52.8, 126.2, 127.1, 128.4, 129.2, 132.8, 134.5, 166.5, 171.6; IR (KBr) 3305, 1673 cm $^{-1}$; HRMS (ESI) $\mathit{m/z}$ calcd for $C_{19}H_{19}N_2O_2SCl$ 397.0748 $(M + Na)^+$, found 397.0751. 7b-B (aS,S) (85% de): colorless oil; $[\alpha]^{25}_{D}$ +21.099 (c 0.1, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 1.52 (d, J = 7.2 Hz, 3H), 2.46 (ddd, *J* = 7.4, 12.8, 12.8 Hz, 1H), 2.65 (ddd, *J* = 1.6, 6.1, 12.8 Hz, 1H), 3.27 (ddd, J = 6.1, 12.8, 12.8, 1H), 3.37 (ddd, J = 1.6, 7.4, 12.8 Hz, 1H), 4.07 (d, J = 16.6 Hz, 1H), 4.89 (d, J = 16.6 Hz, 1H), 5.10 (quin, J = 7.2 Hz, 1H), 7.18–7.35 (m, 6H), 7.52 (dd, J = 0.6, 8.4 Hz, 2H), 7.83 (br-d, 7.6 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.7, 33.2, 34.1, 49.0, 52.3, 126.2, 128.3, 129.0, 130.3, 130.4, 132.7, 134.4, 142.3, 142.9, 166.3, 171.6; IR (neat) 3316, 1680 cm⁻¹; HRMS (ESI) *m/z* calcd for $C_{19}H_{19}N_2O_2SCl 397.0748 (M + Na)^+$, found 397.0747.

2-(9-Chloro-2-oxo-2,3,4,5-tetrahydro-1H-1-benzoazepin-1-yl)-N-[(1S)-1-phenylethyl]acetamide (7c) and the Atropisomers (7c-A and 7c-B). Compounds 7c was prepared from the corresponding acetate (5c) via the carboxylic acid (5c-CO₂H) according to a similar procedure described for the preparation of 7a from 5a.

(9-Chloro-2-oxo-2,3,4,5-tetrahydro-1*H*-1-benzoazepin-1-yl)acetic acid (5c-CO₂H): white solid (90%); mp 143–145 °C; ¹H NMR (400 MHz, CDCl₃) δ 1.88–1.97 (m, 1H), 2.21–2.42 (m, 3H), 2.62 (dd, *J* = 6.5, 13.6 Hz, 1H), 3.57 (ddd, *J* = 7.3, 13.4, 13.6 Hz, 1H), 4.17 (d, *J* = 17.5 Hz, 1H), 4.83 (d, *J* = 17.5 Hz, 1H), 7.14 (dd, *J* = 1.7, 7.5 Hz, 1H), 7.19 (dd, *J* = 7.5, 7.8 Hz, 1H), 7.35 (dd, *J* = 1.7, 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 28.3, 29.8, 32.1, 49.5, 128.0, 128.5, 128.7, 129.0, 138.5, 139.8, 173.0, 174.1; IR (KBr) 2944, 1745, 1624 cm⁻¹; HRMS (ESI) *m/z* calcd for C₁₂H₁₂NO₃Cl 276.0398 (M + Na)⁺, found 276.0397.

2-(9-Chloro-2-oxo-2,3,4,5-tetrahydro-1H-1-benzoazepin-1-yl)-N-[(1S)-1-phenylethyl]acetamide (7c). The mixture (with a ratio of 1:0.8 determined by ¹H NMR) of the diastereomers of 7c-A and 7c-B was obtained as a white foam (yield 87%). The foam was chromatographed on SiO₂ to separate the diastereomers 7c-A (aR,S) and 7**c-B** (a*S*,*S*). 7**c-A** (a*R*,*S*): colorless crystals (isolated yield: 36%); mp $52-54 \,^{\circ}\text{C}; [\alpha]^{24}_{D} - 19.1 (c \, 0.2, \text{MeOH}); {}^{1}\text{H NMR} (400 \,\text{MHz}, \text{CDCl}_3)$ δ 1.47 (d, J = 7.0 Hz, 3H), 1.78–1.84 (m, 1H), 2.09–2.30 (m, 3H), 2.37 (dd, J = 7.3, 10.3 Hz, 1H), 2.84 (ddd, J = 5.9, 10.3, 12.9 Hz, 1H), 3.97 (d, J = 14.9 Hz, 1H), 4.69 (d, J = 14.9 Hz, 1H), 5.08 (quin, J = 7.0 Hz, 1H), 6.73 (br-d, J = 7.6 Hz, 1H), 7.05 (dd, J = 1.2, 7.5 Hz, 1H), 7.15 (t, J = 7.5 Hz, 1H), 7.23–7.35 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 22.0, 28.3, 29.5, 32.2, 48.8, 52.2, 125.9, 126.0, 127.1, 127.6, 128.3, 128.4, 128.4,128.7, 128.9, 138.8, 139.3, 142.8, 167.1, 174.3; IR (KBr) 3306, 2931, 1656 cm⁻¹; HRMS (ESI) m/z calcd for C₂₀H₂₁N₂O₂Cl 379.1184 $(M + Na)^+$, found 379.1181. 7c-B (aS,S): colorless crystals (isolated yield: 31%); mp 78–80 °C; $[\alpha]^{24}_{D}$ –217.4 (c 0.2, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 1.53 (d, J = 7.0 Hz, 3H), 1.88–1.97 (m, 1H), 2.21-2.42 (m, 3H), 2.65 (dd, J = 7.3, 10.5 Hz, 1H), 3.44 (ddd, J = 5.6, 10.5, 12.9 Hz, 1H), 3.99 (d, J = 15.1 Hz, 1H), 4.61 (d, J = 15.1 Hz, 1H), 5.09 (quin, J = 7.0 Hz, 1H), 6.47 (br-d, J = 7.6 Hz, 1H), 7.12 (dd, J = 1.4, 7.5 Hz, 1H), 7.16 (t, J = 7.5 Hz, 1H), 7.22–7.35 (m, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 21.9, 28.4, 29.9, 32.3, 49.0, 52.4, 126.0, 127.2, 127.8, 125.5, 128.8, 128.9, 139.1, 139.5, 142.9, 167.4, 174.3; IR (KBr) 3309, 2942, 1654 cm⁻¹; HRMS (ESI) m/z calcd for C₂₀H₂₁N₂O₂Cl $357.1364 (M + H)^+$, found 357.1370.

2-(5-Methyl-2-oxo-9-phenyl-2,3,4,5-tetrahydro-1*H*-1,5benzodiazepin-1-yl)-*N*-[(1*S*)-1-phenylethyl]acetamide (8a) and the Atropisomers 8a-A and 8a-B. Compound 8a was prepared from the corresponding acetate (6a) via the carboxylic acid (6a-CO₂H) according to a similar procedure described for the preparation of 7a from 5a.

(5-Methyl-2-oxo-9-phenyl-2,3,4,5-tetrahydro-1*H*-1,5-benzodiazepin-1-yl)acetic acid (6a-CO₂H): white solid (94%); mp 179–183 °C; ¹H NMR (400 MHz, CDCl₃) δ 2.57 (dd, *J* = 5.6, 12.9 Hz, 1H), 2.82 (ddd, *J* = 7.3, 12.7, 12.9 Hz, 1H), 2.92 (s, 3H), 3.08 (d, *J* = 18.0 Hz, 1H), 3.14 (dd, *J* = 7.3, 10.5 Hz, 1H), 3.82 (ddd, *J* = 5.6, 10.5, 12.7 Hz, 1H), 4.49 (d, *J* = 18.0 Hz, 1H), 7.25 (dd, *J* = 1.2, 8.0 Hz, 1H), 7.30–7.35 (m, 3H), 7.39–7.47 (m, 3H), 7.50 (dd, *J* = 7.8, 8.0 Hz, 1H), 12.5 (br, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 33.2, 40.6, 50.1, 59.0, 118.8, 128.1, 128.3, 128.5, 129.1, 129.5, 134.5, 136.9, 137.8, 143.1, 169.4, 172.8; IR (KBr) 3491, 1739, 1679 cm⁻¹; HRMS (ESI) *m*/*z* calcd for C₁₈H₁₉N₂O₃ 311.1390 (M + H)⁺, found 311.1387.

2-(5-Methyl-2-oxo-9-phenyl-2,3,4,5-tetrahydro-*1H***-1,5-benzodiazepin-1-yl**)-*N*-[(15)-1-phenylethyl]acetamide (8a). The mixture (with a ratio of 1:0.89 determined by ¹H NMR) of the diastereomers of **8a-A** and **8a-B** was obtained as a colorless form (yield 89%), which was chromatographed on SiO₂ to separate the diastereomers **8a-A** (a*S*,*S*) and **8a-B** (a*R*,*S*). **8a-A** (a*S*,*S*): colorless crystals (isolated yield: 39%); mp 196–198 °C; $[\alpha]^{21}_{D}$ +51.5 (*c* 0.30, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.34 (d, *J* = 6.8 Hz, 3H), 2.22 (s, 3H), 2.47 (dd, *J* = 5.3, 12.9 Hz, 1H), 2.75 (ddd, *J* = 5.3, 7.5, 12.9 Hz, 1H), 2.87 (dd, *J* = 7.5, 10.0 Hz, 1H), 3.08 (d, *J* = 17.3 Hz, 1H), 3.07 (ddd, *J* = 5.3, 10.0, 12.9 Hz, 1H), 4.22 (d, *J* = 17.3 Hz, 1H), 5.09 (quin, *J* = 6.8 Hz, 1H),

7.06 (dd, *J* = 1.2, 8.0 Hz, 1H), 7.21 (dd, *J* = 1.4, 7.8 Hz, 1H), 7.27–7.30 (m, 1H), 7.33-7.46 (m, 10H), 8.33 (br-d, J = 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 20.3, 33.7, 40.3, 47.1, 52.2, 58.6, 118.4, 126.4, 127.1, 128.1, 128.2, 128.4, 128.5, 129.2, 134.4, 137.1, 138.0, 142.7, 144.0, 167.5, 172.4; IR (KBr) 3246, 1676 cm⁻¹; HRMS (ESI) m/z calcd for $C_{26}H_{28}N_3O_2$ 414.2176 (M + H)⁺, found 414.2171. 8a-B (aR,S): colorless crystals (isolated yield: 39%); mp 196–198 °C; $[\alpha]_{D}^{21}$ –46.2 (c 0.30, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 1.47 (d, J = 7.3 Hz, 3H), 2.34 (s, 3H), 2.51 (dd, *J* = 5.3, 12.9 Hz, 1H), 2.82 (ddd, *J* = 5.3, 7.5, 12.9 Hz, 1H), 3.05 (dd, *J* = 7.5, 10.0 Hz, 1H), 3.07 (d, *J* = 17.3 Hz, 1H), 3.72 (ddd, *J* = 5.3, 10.0, 12.9 Hz, 1H), 4.31 (d, J = 17.3 Hz, 1H), 5.08 (quin, J = 7.3 Hz, 1H), 6.99-7.01(m, 3H), 7.17-7.22 (m, 4H), 7.33-7.45 (m, 6H), 8.10 (br-d, J = 7.8 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃) δ 21.2, 33.8, 40.6, 47.4, 51.5, 59.3, 118.4, 126.1, 126.9, 127.1, 128.1, 128.2, 128.3, 128.4, 129.2, 134.0, 137.1, 138.0, 143.1, 144.0, 167.3, 172.5. IR (KBr) 3275, 1678 cm⁻¹; HRMS (ESI) m/z calcd for C₂₆H₂₈N₃O₂ 414.2176 (M + H)⁺, found 414.2171.

Single-Crystal X-ray Analysis. All measurements were made on a RIGAKU RAXIS RAPID imaging plate area detector with graphitemonochromated Cu K α radiation. The data were collected at a temperature of -100 °C. The structure was solved by direct method SIR92 and expanded using Fourier techniques. The non-hydrogen atoms were refined anisotropically. All calculations were performed using Crystal Structure (Crystal Structure 3.8) crystallographic software package. Typical crystal data are as follows.

Cryatal Data for 5b. C₁₂H₁₂O₃NCIS: mp 93–96 °C; M_r = 285.74, Cu Kα (λ = 1.54187 Å), monoclinic, P_{2_1}/n , colorless prism 0.60 × 0.50 × 0.30 mm, crystal dimensions *a* = 9.31754(17) Å, *b* = 11.0378(2) Å, *c* = 24.8795(5) Å, α = 90°, β = 91.2800(7)°, γ = 90°, T = 173 K, Z = 8, V = 2558.10(8) Å³, D_{calc} = 1.484 gcm⁻³, μ Cu Kα = 41.854 cm⁻¹, F_{000} = 1184.00, GOF = 1.227, R_{int} = 0.058, R_1 = 0.0468, w R_2 = 0.1304.

Crystal Data for 7a-A (a*R*,*S***)**. C₂₀H₂₂O₂N₃Cl: mp 173–175 °C, $M_r = 371.87$, Cu Kα ($\lambda = 1.54187$ Å), orthorhombic, P2₁2₁2₁, colorless prism 0.50 × 0.40 × 0.20 mm, crystal dimensions a = 8.18197(15) Å, b = 12.6373(2) Å, c = 17.1203(3) Å, $\alpha = 90^{\circ}$, $\beta = 90^{\circ}$, $\gamma = 90^{\circ}$, T = 173 K, Z = 4, V = 1875.71(6) Å³, $D_{calc} = 1.315$ gcm⁻³, μ Cu K $\alpha = 19.565$ cm⁻¹, $F_{000} = 784.00$, GOF = 0.648, $R_{int} = 0.136$, $R_1 = 0.0471$, w $R_2 = 0.1054$, Flack parameter = 0.04(6).

Crystal Data for 7a-B (a*S,***S**). $C_{20}H_{22}O_2N_3Cl: mp 124-127 °C, M_r = 371.87, Cu Kα (<math>\lambda = 1.54187$ Å), orthorhombic, $P2_12_12_1$, colorless prism 0.25 × 0.20 × 0.10 mm, crystal dimensions a = 8.18197(15) Å, b = 8.69418(16) Å, c = 26.3682(5) Å, $\alpha = 90^\circ$, $\beta = 90^\circ$, $\gamma = 90^\circ$, T = 173 K, Z = 4, V = 1875.71(6) Å³, $D_{calc} = 1.317$ gcm⁻³, μ Cu K $\alpha = 19.583$ cm⁻¹, $F_{000} = 784.00$, GOF = 0.737, $R_{int} = 0.040$, $R_1 = 0.0286$, w $R_2 = 0.0660$, Flack parameter = 0.003(14).

Crystal Data for 7b-A (a*R*,*S***).** $C_{19}H_{19}O_2N_2CIS: mp 142-144 \,^{\circ}C,$ $M_r = 374.88$, Cu K α ($\lambda = 1.54187 \,^{\circ}A$), monoclinic, $P2_1$, colorless prism $0.60 \times 0.40 \times 0.40$ mm, crystal dimensions $a = 8.3785(4) \,^{\circ}A$, $b = 12.3205(4) \,^{\circ}A$, $c = 8.5550(3) \,^{\circ}A$, $\alpha = 90^{\circ}$, $\beta = 90.842(3)^{\circ}$, $\gamma = 90^{\circ}$, T = 173K, Z = 2, $V = 1875.71(6) \,^{\circ}A^3$, $D_{calc} = 1.3410 \, \text{gcm}^{-3}$, μ Cu K $\alpha = 31.449 \, \text{cm}^{-1}$, $F_{000} = 392.00$, GOF = 0.737, $R_{int} = 0.045$, $R_1 = 0.0385$, $wR_2 = 0.0906$, Flack parameter = 0.00(2)

Stereochemical (Thermodynamic) Stability of Diastereomeric Atropisomers (A/B of 7a–c and 8a). The ΔG^{\ddagger} value was determined on the basis of the time-dependent conversion rate (% de) estimated from chiral or nonchiral HPLC analysis of a solution of the diastereomeric atropisomers (A and B) after the compounds were allowed to stand at designated temperatures.¹⁷

ASSOCIATED CONTENT

Supporting Information. General experimental procedure, ¹H NMR and ¹³C NMR spectra for new compounds, figures of thermal isomerization rate of enantiomers of 7**a**-**c** and **8a**, X-ray data (CIF) for compounds **5b**, 7**a**-**A**, 7**a**-**B**, and 7**b**-**A**, and ¹H NMR spectra of the temperature dependent analysis for **4c**. This material is available free of charge via the Internet at http://pubs.acs.org.

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(13) Temperature dependent ¹H NMR analysis was conducted for compound 4c (X = CH₂, Y = H) to reveal that 4c exists as a racemate of the atropisomers at -40 °C in CDCl₃. The activation free-energy barrier to rotation (ΔG^{\dagger}) value estimated from this analysis was ca. 56 kJ/mol. See the Supporting Information.

(14) Note that the definition of the axial chirality (a*S* and a*R*) leads to reversed stereochemical designations between 5, 7 (Y = Cl) and 6, 8 (Y = Ph) according to the priority rule. The terms a*S* and a*R* (chiral axis nomenclature) correspond to *P* and *M* (helix nomenclature), respectively.

(15) The absolute stereochemistry was determined based on the Flack parameter (Cu K α radiation was used for the measurement).

(16) Note that the numbering H^2 of 7b corresponds to H^4 of 7a/7c according to the numbering rule.

(17) For determination of the ΔG^{\ddagger} value, see: Petit, M.; Lapierre, A. J. B.; Curran, D. P. J. Am. Chem. Soc. **2005**, 127, 14994–14995.

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